

## Host Factors Involved in the Growth of Microvirid Phage $\alpha 3$

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Host factors involved in the growth of microvirid phage  $\alpha 3$  were determined using various replication mutants of *Escherichia coli*. The viral multiplication was dependent on functional products of *dnaE*, *dnaF*(*nrda*), *dnaG*, and *dnaZ* genes. Host functions directed by *dnaA*, so-called *dnaH*, *dnaI*, and *dnaP* genes were dispensable for the viral growth. In contrast with  $\phi$ X174 and G4,  $\alpha 3$  would grow sufficiently in *dnaB* and *dnaC*(*D*) mutants. The viral growth was not significantly affected by host *polA*<sup>ts</sup>, *seg*, and *groPC* mutations.

## Introduction

Recently, considerable diversities have been detected among microvirid (isometric) phages, concerning host factor dependence, immunological relationship, and host range. Thus  $\phi$ X174 members (*e. g.*  $\phi$ A, S13, G6) infect *E. coli* C and require host *dnaB* and *dnaC*(*D*) functions for their growth [1–3]. These two host functions are essential for immunologically unrelated phage G4 [4]. Moreover, G13 and G14, which can infect *E. coli* B as well, rely on *dnaB* and *dnaC*(*D*) activities [5]. On the other hand, neither *dnaB* nor *dnaC*(*D*) gene product is required for multiplication of K12-specific group (including  $\phi$ K and St-1) [6] which is immunologically remote from  $\phi$ X174. Assignment of host factors required for these phages is important for elucidation of mechanisms of viral replication and evolution. This report describes the host factor reliance of  $\alpha 3$  [7] which is infective to *E. coli* C and B and immunologically somewhat related to St-1 but not to  $\phi$ X174.

## Materials and Methods

*E. coli* C-N27 *polA4113* [8] was obtained from Dr. T. Okazaki. The sources of other *E. coli* strains

used were previously described [1–6, 9–12]. Unless otherwise specified, bacteria were grown in a nutrient broth, at 30 °C, with shaking. Phage  $\alpha 3$ , originally provided by Dr. D. E. Bradley, was propagated on *E. coli* C and partially purified by differential centrifugation. Single-stranded viral DNA (SS) and double-stranded replicative-form DNA (RF) of  $\alpha 3$  were prepared as previously described [13]. Infection experiments with  $\alpha 3$  phage were performed as described for  $\phi$ K[6]. Bacterial strains resistant to  $\alpha 3$  were transfected with  $\alpha 3$  SS or RF, after Ca<sup>2+</sup>-treatment [1]. The free phage titer was determined using *E. coli* C as the indicator.

## Results and Discussion

Effects of various host mutations on the growth of  $\alpha 3$  are summarized in Table I. Like other microvirid phages,  $\alpha 3$  could grow sufficiently in *dnaA* cells. Moreover, the viral growth proceeded normally in *dnaB* and *dnaC*(*D*) mutants, at 43 °C. On the other hand, multiplication of  $\phi$ X174,  $\phi$ A,

Table I. Growth of  $\alpha 3$  in replication mutants of *E. coli*.

Strain	Phage yield		
	43 °	33 °	43 °/33 °
C <i>dna</i> <sup>+</sup>	(2.4 × 10 <sup>6</sup> ) <sup>a</sup>	2.3 × 10 <sup>6</sup>	1.0
C2307 <i>dnaA</i>	(4.2 × 10 <sup>7</sup> ) <sup>a</sup>	3.2 × 10 <sup>7</sup>	1.3
LD312 <i>dnaB</i>	1.9 × 10 <sup>7</sup>	6.6 × 10 <sup>6</sup>	2.9
LD332 <i>dnaC</i> ( <i>D</i> )	(1.7 × 10 <sup>6</sup> ) <sup>a</sup>	1.6 × 10 <sup>6</sup>	1.1
LD301 <i>dnaE</i>	1.0 × 10 <sup>4</sup>	2.5 × 10 <sup>5</sup>	4.0 × 10 <sup>−2</sup>
JG42 <i>dnaF</i>	(1.1 × 10 <sup>5</sup> ) <sup>b</sup>	(4.7 × 10 <sup>6</sup> ) <sup>b</sup>	2.3 × 10 <sup>−2</sup>
C2309 <i>dnaG</i>	8.0 × 10 <sup>3</sup>	7.0 × 10 <sup>3</sup>	1.1 × 10 <sup>−2</sup>
HF4704S <i>dnaH</i>	(7.3 × 10 <sup>5</sup> ) <sup>a</sup>	7.2 × 10 <sup>5</sup>	1.0
WM301-208 <i>dnaI</i>	(1.9 × 10 <sup>7</sup> ) <sup>a</sup>	1.3 × 10 <sup>7</sup>	1.5
KM107 <i>dnaP</i>	(7.7 × 10 <sup>6</sup> ) <sup>c</sup>	9.1 × 10 <sup>6</sup>	8.5 × 10 <sup>−1</sup>
AX727 <i>dnaZ</i>	(1.6 × 10 <sup>3</sup> ) <sup>b</sup>	(1.4 × 10 <sup>6</sup> ) <sup>b</sup>	1.1 × 10 <sup>−3</sup>
C727 <i>dnaZ</i>	1.5 × 10 <sup>4</sup>	2.2 × 10 <sup>6</sup>	6.8 × 10 <sup>−3</sup>
C-N27 <i>polA 4113</i>	6.9 × 10 <sup>6</sup>	2.4 × 10 <sup>7</sup>	2.9 × 10 <sup>−1</sup>
BT4113 <i>polA</i> <sup>ts</sup>	(3.3 × 10 <sup>5</sup> ) <sup>b</sup>	(5.2 × 10 <sup>5</sup> ) <sup>b</sup>	4.4 × 10 <sup>−1</sup>
KS268 <i>ligts7</i>	(1.8 × 10 <sup>4</sup> ) <sup>b</sup>	(3.0 × 10 <sup>6</sup> ) <sup>b</sup>	6.0 × 10 <sup>−3</sup>
BW2001 <i>xth-11</i>	(2.0 × 10 <sup>5</sup> ) <sup>b</sup>	(2.6 × 10 <sup>6</sup> ) <sup>b</sup>	7.7 × 10 <sup>−2</sup>
PB213 <i>seg</i> <sup>+</sup>	(3.4 × 10 <sup>3</sup> ) <sup>b</sup>	(6.6 × 10 <sup>3</sup> ) <sup>b</sup>	5.2 × 10 <sup>−1</sup>
PB1022 <i>seg-2</i>	(1.1 × 10 <sup>3</sup> ) <sup>b</sup>	(3.8 × 10 <sup>3</sup> ) <sup>b</sup>	2.9 × 10 <sup>−1</sup>
PB1022 <i>seg-2</i>	(2.3 × 10 <sup>5</sup> ) <sup>d</sup>	(3.7 × 10 <sup>4</sup> ) <sup>d</sup>	6.2
C600 <i>gro</i> <sup>+</sup>	(3.4 × 10 <sup>4</sup> ) <sup>b</sup> *	(4.1 × 10 <sup>4</sup> ) <sup>b</sup>	8.3 × 10 <sup>−1</sup>
MF634 <i>groPC</i> 259	(1.3 × 10 <sup>4</sup> ) <sup>b</sup> *	(2.6 × 10 <sup>4</sup> ) <sup>b</sup>	5.0 × 10 <sup>−1</sup>
C600 <i>groPC</i> 756	(9.9 × 10 <sup>3</sup> ) <sup>b</sup> *	(7.1 × 10 <sup>4</sup> ) <sup>b</sup>	1.4 × 10 <sup>−1</sup>
C600 <i>groPC</i> 756	(1.9 × 10 <sup>5</sup> ) <sup>d</sup> *	(5.3 × 10 <sup>3</sup> ) <sup>d</sup>	3.6 × 10 <sup>−1</sup>

<sup>a</sup> Cells were grown for 60 min at 43 °C before infection.

<sup>b</sup> Phage yield was determined by transfection of SS DNA to the Ca<sup>2+</sup>-treated bacteria.

<sup>c</sup> Cells were grown for 100 min at 43 °C prior to infection.

<sup>d</sup> Phage yield was determined by transfection of RF DNA to the Ca<sup>2+</sup>-treated cells.

\* Incubation temperature was 43.5 °C.

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S13, and G6 was clearly restricted in these hosts, at the high temperature. In *dnaE*, *dnaF*(*nrdA*), and *dnaG* mutants, growth of  $\alpha 3$  was thermosensitive, indicating involvement of host DNA polymerase III, ribonucleotide reductase, and primase in the viral replication. Yield of  $\alpha 3$  was not reduced at 43 °C in HF4704S "*dnaH*" which was recently shown to carry double mutations in *dnaA* gene and in some step for utilization of exogenous thymine [14]. (Among enzymes participated in thymine utilization, thymidine phosphorylase and purine nucleoside phosphorylase were not particularly defective in HF4704S strain.) At 43 °C, replication of  $\alpha 3$  was not significantly affected in strains WM301-208 *dnaI* and KM107 *dnaP*. Furthermore, growth of  $\varphi$ X174 was not thermosensitive in the *dnaP* strain (unpublished observation). In contrast, multiplication of  $\alpha 3$  was distinctly thermosensitive in AX727 *dnaZ* mutant transfected with  $\alpha 3$  SS, as well as in C727 *dnaZ* cells infected with intact  $\alpha 3$  phage. Functional product of *dnaZ* gene is essential for all microvirid phages thus far tested. It must be noted here that host functions directed by *dnaE*, *dnaF*(*nrdA*), *dnaG*, and *dnaZ* genes are essential for  $\lambda$  phage as well.

Although multiplication of  $\alpha 3$  was only slightly affected by *polA*<sup>ts</sup> mutation, the phage yield was markedly reduced at 43 °C in strain KS268 *ligts7*, indicating participation of host DNA ligase in the viral growth. In strain BW2001 *xth-11*, growth of  $\alpha 3$  was considerably reduced at 43 °C. However, whether exonuclease III, product of the *xth* gene, is directly involved in  $\alpha 3$  replication process (*e. g.* removal of primer RNA) or not is presently un-

known. Replication of  $\alpha 3$  was not significantly affected at 43 °C in PB1022 *seg-2* cells, whereas growth of  $\lambda$  phage was, as reported by Jamieson and Bergquist [15], abortive in this mutant at 42 °C–43 °C (data not shown). In contrast with  $\lambda$ ,  $\alpha 3$  could grow normally in *groPC* mutants, at 37 °C. Yield of  $\alpha 3$  in strains MF634 *groPC*259 and C600 *groPC*756 was not particularly reduced even at 43.5 °C, as compared with the yield at 33 °C. Unlike  $\lambda$  phage,  $\alpha 3$  requires host functions specified by *rep* gene (data not shown).

Host range of  $\alpha 3$  is similar to that of G13 and G14: these phages can infect *E. coli* BB and BB5 but not to BB2, BB1 BB20, BB7, BB9, BB4 and BB12 (unpublished observation). Host factor requirement of  $\alpha 3$  nevertheless differs from that of G13 and G14, and closely resembles that of K12-specific phages St-1 and  $\varphi$ K [6]. Moreover,  $\varphi$ Kh-1 (a host range mutant of  $\varphi$ K) [2] can infect *E. coli* K12, C, BB, BB5, BB1, and BB7, whereas host factor reliance of this phage is essentially similar to St-1,  $\varphi$ K, and  $\alpha 3$ . These results are consistent with the fact that  $\alpha 3$  is immunologically related to St-1 group [7] but quite different G13 and G14.

Dispensability of *dnaB* and *dnaC*(*D*) functions for  $\alpha 3$  predicts that this phage, like G4 [16], may have a unique origin of synthesis of complementary (minus) DNA strand. Furthermore, in synthesis of viral (plus) DNA strand,  $\alpha 3$  system is by far simpler than G4 which, like  $\varphi$ X174, requires *dnaB* and *dnaC*(*D*) genes products for this reaction [4]. Determination of nucleotide sequence of the origin of plus strand synthesis is essential for characterization of this unique replication system.

- [1] A. Taketo, *Molec. Gen. Genet.* **122**, 15 (1973).
- [2] A. Taketo, *J. Gen. Appl. Microbiol.* **23**, 29 (1977).
- [3] A. Taketo, *J. Gen. Appl. Microbiol.* **23**, 85 (1977).
- [4] K. Kodaira and A. Taketo, *J. Biochem.* **83**, 971 (1978).
- [5] A. Taketo and K. Kodaira, *Biochim. Biophys. Acta*, in press.
- [6] A. Taketo, *Molec. Gen. Genet.* **148**, 25 (1976).
- [7] D. E. Bradley, *Canad. J. Microbiol.* **16**, 965 (1970).
- [8] T. Ogawa, S. Hirose, T. Okazaki, and R. Okazaki, *J. Molec. Biol.* **112**, 121 (1977).
- [9] A. Taketo, *Molec. Gen. Genet.* **139**, 285 (1975).
- [10] K. Kodaira and A. Taketo, *Biochim. Biophys. Acta* **476**, 149 (1977).
- [11] A. Taketo and K. Kodaira, *Molec. Gen. Genet.* **162**, 151 (1978).
- [12] A. Taketo, *Z. Naturforsch.* **32c**, 429 (1977).
- [13] A. Taketo, *J. Gen. Appl. Microbiol.* **24**, 51 (1978).
- [14] P. L. Derstine and L. B. Dumas, *J. Bacteriol.* **128**, 801 (1976).
- [15] A. F. Jamieson and P. L. Bergquist, *Molec. Gen. Genet.* **150**, 171 (1977).
- [16] J. C. Fiddes, B. G. Barell, and G. N. Godson, *Proc. Nat. Acad. Sci. USA* **75**, 1081 (1978).